



CHALMERS
UNIVERSITY OF TECHNOLOGY

Draft genome sequence of bacillus coagulans ma-13, a thermophilic lactic acid producer from lignocellulose

Downloaded from: <https://research.chalmers.se>, 2024-04-16 07:57 UTC

Citation for the original published paper (version of record):

Aulitto, M., Fusco, S., Franzén, C. et al (2019). Draft genome sequence of bacillus coagulans ma-13, a thermophilic lactic acid producer from lignocellulose. Microbiology Resource Announcements, 8(23).
<http://dx.doi.org/10.1128/MRA.00341-19>

N.B. When citing this work, cite the original published paper.



Draft Genome Sequence of *Bacillus coagulans* MA-13, a Thermophilic Lactic Acid Producer from Lignocellulose

 Martina Aulitto,^{a,b}  Salvatore Fusco,^{a,b}  Carl Johan Franzén,^b  Andrea Strazzulli,^{a,c}  Marco Moracci,^{a,c}
 Simonetta Bartolucci,^a  Patrizia Contursi^{a,c}

^aDepartment of Biology, University of Naples Federico II, Naples, Italy

^bDivision of Industrial Biotechnology, Department of Biology and Biological Engineering, Chalmers University of Technology, Gothenburg, Sweden

^cTask Force on Microbiome Studies, University of Naples Federico II, Naples, Italy

ABSTRACT *Bacillus coagulans* MA-13 is an efficient lactic acid producer which withstands high concentrations of the growth inhibitors formed during the pretreatment of lignocellulosic feedstock. This draft genome sequence is expected to pave the way toward the understanding of mechanisms responsible for the robustness of MA-13 during simultaneous saccharification and fermentation.

Bacillus coagulans MA-13 is a Gram-positive spore-forming moderately thermophilic facultatively anaerobic bacterium isolated from processed bean waste (1). MA-13 ferments lignocellulose-derived hexoses to lactic acid (LA); therefore, it is a suitable candidate for the conversion of lignocellulose to LA, which is a building block for the production of polylactic acid (PLA), i.e., a biodegradable bioplastic (2). Recently, MA-13 was used for the conversion of steam-exploded wheat straw to LA in simultaneous saccharification and fermentation (SSF) (3, 4). The preexposure to the inhibitor-rich lignocellulosic hydrolysate (5) led to a physiological adaptation of MA-13, which was reflected in an improved fermentation performance during SSF, thus resulting in a more cost-effective process (4).

The strain MA-13 was isolated and cultivated as previously described (1) before genomic DNA was extracted using the LETS (lithium, EDTA, Tris, and SDS) buffer method, followed by phenol extraction (6). The sequencing of the whole genome was performed using the Illumina NextSeq platform at Genomix4life S.R.L. (Salerno, Italy) with paired-end indexed libraries prepared using a Nextera XT kit (Illumina, Inc.). The reads (151 nucleotides [nt]) were *de novo* assembled using the SPAdes genome assembler version 3.9.0 on BaseSpace (7, 8). A total of 11,245,275 paired-end reads with an average length of 150 base pairs (bp) were assembled into 1,653 contigs (N_{50} length of 51,225 nt, N_{90} length of 4,278 nt), with the largest contig being 145,076 nt long. The draft genome consists of 3,237,270 bp with a GC content of 47.11%.

Functional annotation of contigs was carried out using the comprehensive bioinformatics tool Blast2GO version 5.2.5 (9, 10). A total of 3,336 open reading frames (ORFs) were identified, 3,268 of which were predicted as genes. A further annotation analysis was carried out with Rapid Annotations using Subsystems Technology (RAST) software (myRAST version 36) (11). Default parameters were used for all software unless otherwise specified. There were 2,355 gene ontology (GO) terms, 468 of which were assigned to the category of biological processes, including all necessary genes for the glycolysis (Embden-Meyerhof-Parnas) and the tricarboxylic acid cycle. Moreover, one D-lactate and three L-lactate dehydrogenase genes were identified, which can account for the superior fermentation performance of MA-13 (1, 4). The tolerance toward lignocellulose-derived inhibitors can be traced back to the presence in the MA-13 genome of genes encoding enzymes putatively involved in

Citation Aulitto M, Fusco S, Franzén CJ, Strazzulli A, Moracci M, Bartolucci S, Contursi P. 2019. Draft genome sequence of *Bacillus coagulans* MA-13, a thermophilic lactic acid producer from lignocellulose. Microbiol Resour Announc 8:e00341-19. <https://doi.org/10.1128/MRA.00341-19>.

Editor David A. Baltrus, University of Arizona

Copyright © 2019 Aulitto et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Patrizia Contursi, contursi@unina.it.

M.A. and S.F. contributed equally to this work.

Received 5 April 2019

Accepted 10 May 2019

Published 6 June 2019

detoxification pathways, i.e., four aldehyde dehydrogenases, three short-chain dehydrogenases, two alcohol dehydrogenases, and one zinc-dependent alcohol dehydrogenase, which are potentially associated with detoxification reactions (12–14). Besides LA metabolism, MA-13 possesses genes required for the production of value-added chemicals, such as acetoin, butanediol, and polyhydroxybutyrate (i.e., a biodegradable plastic). The presence of genes encoding bacteriocins is related to the production of antimicrobial molecules (15–17) suitable to avoid competition with other bacteria in nonsterile open fermentation. As shown for other *B. coagulans* strains (18–21), genes associated with the defense mechanism toward foreign genetic elements, i.e., the clusters of regularly interspaced short palindromic repeat (CRISPR)-cas systems (22), were identified using CRISPRFinder version 1.3 (23).

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [SMSP00000000](https://www.ncbi.nlm.nih.gov/Traces/study/?acc=PRJNA526660). The version described in this paper is version SMSP01000000. The raw reads have been deposited in the SRA under the accession number [PRJNA526660](https://www.ncbi.nlm.nih.gov/Traces/study/?acc=PRJNA526660) and are also available at <https://www.ncbi.nlm.nih.gov/Traces/study/?acc=PRJNA526660>.

ACKNOWLEDGMENTS

This research was carried out under the Programme STAR and financially supported by UniNA and Compagnia di San Paolo (grant number 16-CSP-UNINA-007). The funding bodies had no influence on the design of the study and were not involved in the collection, analysis, or interpretation of data or in the writing of the manuscript.

All authors contributed to the conception and planning of the study. M.A. and S.F. performed the experiments and drafted the manuscript. M.A., S.F., and A.S. carried out *in silico* analyses of enzymes potentially involved in the detoxification reaction as well as polysaccharide and lactate metabolism. M.M., S.B., C.J.F., and P.C. supervised the experimental work and reviewed the manuscript. All the authors read and approved the final version of the manuscript.

REFERENCES

- Aulitto M, Fusco S, Bartolucci S, Franzén CJ, Contursi P. 2017. *Bacillus coagulans* MA-13: a promising thermophilic and cellulolytic strain for the production of lactic acid from lignocellulosic hydrolysate. *Biotechnol Biofuels* 10:210. <https://doi.org/10.1186/s13068-017-0896-8>.
- Komesu A, de Oliveira JAR, da Silva Martins LH, Maciel MRW, Maciel Filho R. 2017. Lactic acid production to purification: a review. *BioResources* 12:4364–4383. <https://doi.org/10.15376/biores.12.2.Komesu>.
- Aulitto M, Fusco S, Fiorentino G, Limauro D, Pedone E, Bartolucci S, Contursi P. 2017. *Thermus thermophilus* as source of thermozymes for biotechnological applications: homologous expression and biochemical characterization of an α -galactosidase. *Microb Cell Fact* 13:16:28. <https://doi.org/10.1186/s12934-017-0638-4>.
- Aulitto M, Fusco S, Nickel DB, Bartolucci S, Contursi P, Franzén CJ. 2019. Seed culture pre-adaptation of *Bacillus coagulans* MA-13 improves lactic acid production in simultaneous saccharification and fermentation. *Biotechnol Biofuels* 12:45. <https://doi.org/10.1186/s13068-019-1382-2>.
- Wang R, Unrean P, Franzén CJ. 2016. Model-based optimization and scale-up of multi-feed simultaneous saccharification and co-fermentation of steam pre-treated lignocellulose enables high gravity ethanol production. *Biotechnol Biofuels* 9:88. <https://doi.org/10.1186/s13068-016-0500-7>.
- Fusco S, She Q, Fiorentino G, Bartolucci S, Contursi P. 2015. Unravelling the role of the F55 regulator in the transition from lysogeny to UV induction of *Sulfolobus* spindle-shaped virus 1. *J Virol* 89:6453–6461. <https://doi.org/10.1128/JVI.00363-15>.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20:714–737. <https://doi.org/10.1089/cmb.2013.0084>.
- Kulikov AS, Prjibelski AD, Tesler G, Vyahhi N, Sirotkin AV, Pham S, Dvorkin M, Pevzner PA, Bankevich A, Nikolenko SI, Pyshkin AV, Nurk S, Gurevich AA, Antipov D, Alekseyev MA, Lesin VM. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Conesa A, Götz S, García-Gómez JM, Terol J, Talón M, Robles M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 21:3674–3676. <https://doi.org/10.1093/bioinformatics/bti610>.
- Conesa A, Götz S. 2008. Blast2GO: a comprehensive suite for functional analysis in plant genomics. *Int J Plant Genomics* 2008:619832.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Wang X, Yomano LP, Lee JY, York SW, Zheng H, Mullinnix MT, Shanmugam K, Ingram LO. 2013. Engineering furfural tolerance in *Escherichia coli* improves the fermentation of lignocellulosic sugars into renewable chemicals. *Proc Natl Acad Sci U S A* 110:4021–4026. <https://doi.org/10.1073/pnas.1217958110>.
- Peng L, Song L, Sun L, Cai Y, Wang L, Yu B. 2015. Genome sequence of *Bacillus coagulans* P38, an efficient polymer-grade L-lactate producer from cellulosic substrates. *Genome Announc* 3:e00495-15. <https://doi.org/10.1128/genomeA.00495-15>.
- Kang C, Hayes R, Sanchez EJ, Webb BN, Li Q, Hooper T, Nissen MS, Xun L. 2012. Furfural reduction mechanism of a zinc-dependent alcohol dehydrogenase from *Cupriavidus necator* JMP134. *Mol Microbiol* 83: 85–95. <https://doi.org/10.1111/j.1365-2958.2011.07914.x>.
- Notomista E, Falanga A, Fusco S, Pirone L, Zanfardino A, Galdiero S, Varcamonti M, Pedone E, Contursi P. 2015. The identification of a novel *Sulfolobus islandicus* CAMP-like peptide points to archaeal microorganisms as cell factories for the production of antimicrobial molecules. *Microb Cell Fact* 14:126. <https://doi.org/10.1186/s12934-015-0302-9>.
- Gaglione R, Pirone L, Farina B, Fusco S, Smaledone G, Aulitto M, Dell'Olmo

- E, Roscetto E, Del Gatto A, Fattorusso R, Notomista E, Zaccaro L, Arciello A, Pedone E, Contursi P. 2017. Insights into the anticancer properties of the first antimicrobial peptide from Archaea. *Biochim Biophys Acta Gen Subj* 1861:2155–2164. <https://doi.org/10.1016/j.bbagen.2017.06.009>.
17. Roscetto E, Contursi P, Vollaro A, Fusco S, Notomista E, Catania MR. 2018. Antifungal and anti-biofilm activity of the first cryptic antimicrobial peptide from an archaeal protein against *Candida* spp. clinical isolates. *Sci Rep* 8:17570. <https://doi.org/10.1038/s41598-018-35530-0>.
18. Orrù L, Salvetti E, Cattivelli L, Lamontanara A, Michelotti V, Capozzi V, Spano G, Keller D, Cash H, Martina A, Torriani S, Felis GE. 2014. Draft genome sequence of *Bacillus coagulans* GBI-30, 6086, a widely used spore-forming probiotic strain. *Genome Announc* 2:e01080-14. <https://doi.org/10.1128/genomeA.01080-14>.
19. Upadrasta A, Pitta S, Madempudi RS. 2016. Draft genome sequence of the spore-forming probiotic strain *Bacillus coagulans* Unique IS-2. *Genome Announc* 4:e00225-16. <https://doi.org/10.1128/genomeA.00225-16>.
20. Zheng Z, Jiang T, Lin X, Zhou J, Ouyang J. 2015. Draft genome sequence of *Bacillus coagulans* NL01, a wonderful L-lactic acid producer. *Genome Announc* 3:e00635-15. <https://doi.org/10.1128/genomeA.00635-15>.
21. Su F, Tao F, Tang H, Xu P. 2012. Genome sequence of the thermophile *Bacillus coagulans* Hammer, the type strain of the species. *J Bacteriol* 194:6294–6295. <https://doi.org/10.1128/JB.01380-12>.
22. Fusco S, Liguori R, Limauro D, Bartolucci S, She Q, Contursi P. 2015. Transcriptome analysis of *Sulfolobus solfataricus* infected with two related fuselloviruses reveals novel insights into the regulation of CRISPR-Cas system. *Biochimie* 118:322–332. <https://doi.org/10.1016/j.biochi.2015.04.006>.
23. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <https://doi.org/10.1093/nar/gkm360>.