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Potential infection of grazing cattle via contaminated water: a theoretical modelling approach

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Wastewater discharge and agricultural activities may pose microbial risks to natural water sources. The impact of different sources can be assessed by water quality modelling. The aim of this study was to use hydrological and hydrodynamic models to illustrate the risk of exposing grazing animals to faecal pollutants in natural water sources, using three zoonotic faecal pathogens as model microbes and fictitious pastures in Sweden as examples. Microbial contamination by manure from fertilisation and grazing was modelled by use of a hydrological model (HYPE) and a hydrodynamic model (MIKE 3 FM), and microbial contamination from human wastewater was modelled by application of both models in a backwards process. The faecal pathogens Salmonella spp., verotoxin-producing Escherichia coli O157:H7 (VTEC) and Cryptosporidium parvum were chosen as model organisms. The pathogen loads on arable land and pastures were estimated based on pathogen concentration in cattle faeces, herd prevalence and within-herd prevalence. Contamination from human wastewater discharge was simulated by estimating the number of pathogens required from a fictitious wastewater discharge to reach a concentration high enough to cause infection in cattle using the points on the fictitious pastures as their primary source of drinking water. In the scenarios for pathogens from animal sources, none of the simulated concentrations of salmonella exceeded the concentrations needed to infect adult cattle. For VTEC, most of the simulated concentrations exceeded the concentration needed to infect calves. For C. parvum, all the simulated concentrations exceeded the concentration needed to infect calves. The pathogen loads needed at the release points for human wastewater to achieve infectious doses for cattle were mostly above the potential loads of salmonella and VTEC estimated to be present in a 24-h overflow from a medium-size Swedish wastewater treatment plant, while the required pathogen loads of C. parvum at the release points were below the potential loads of C. parvum in a 24-h wastewater overflow. Most estimates in this study assume a worst-case scenario. Controlling zoonotic infections at herd level prevents environmental contamination and subsequent human exposure. The potential for infection of grazing animals with faecal pathogens has implications for keeping animals on pastures with access to natural water sources. As the infectious dose for most pathogens is more easily reached for calves than for adult animals, and young calves are also the main shedders of C. parvum, keeping young calves on pastures adjacent to natural water sources is best avoided.

Keywords: Salmonella, Cryptosporidium, bovine, hydrological, hydrodynamic

Implications

We demonstrate that water quality modelling should also be used to assess microbial risks for animals, and that combining hydrological and hydrodynamic models and implementing backwards modelling is useful for this purpose. The importance of on-farm control of diseases for animal and public health is illustrated, as well as the importance of human wastewater treatment. To obtain valid model

outputs, detailed input data from surveillance of animal and human diseases are needed.

Introduction

Wastewater discharges, for example from municipal wastewater treatment plants, and agricultural activities, such as application of manure on arable land and grazing animals on pastures, may pose microbial risks to water sources used for drinking water supply and recreation. However, in addition to providing a potential source for microbial contamination,

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grazing animals may be recipients of microbes via contaminated water. Further, after being infected from microbes in water, grazing animals may provide subsequent sources of microbial contamination, such as seen in, for example, outbreaks of salmonella (Clark *et al.*, 2004; Vanselow *et al.*, 2007; Lahti *et al.*, 2010).

Grazing animals are needed for maintaining certain nature reserves and promote biodiversity, and natural water sources often form part of these ecosystems. Pastureland outside nature reserves may also contain natural water sources that are utilised by the grazing animals regardless of the provision of clean drinking water in other locations on the pasture. Restricting the animals' access to natural water sources may be costly as well as impractical.

There is a need to appropriately assess the risk of exposing grazing animals to microbes that could potentially spread via water in concentrations corresponding to infectious doses. Microbes from manure application on arable land, from faeces of grazing animals and from human wastewater can reach the animals' water sources by transport through soil, but mainly via surface runoff, from the immediate vicinity or via streams, rivers and lakes. The complexity of the water transport processes and lack of quantitative input data hamper any direct assessment of this risk. However, the relative impact of different sources can be assessed by simulating the fate and transport of microbial pollutants in different catchments using water quality modelling (Oliver et al., 2016). Hydrological (also called catchment/watershedscale) models have been widely applied to estimate the impacts of diffuse faecal sources in agricultural catchments worldwide (Jamieson et al., 2004: Bradford et al., 2013: Cho et al., 2016). Hydrodynamic models have also been applied to estimate the impacts of particularly point faecal sources on water bodies (e.g. lakes, rivers, estuaries and coastal zones) used for drinking water production, recreation and shellfish growing in different countries (De Brauwere et al., 2014b; Dienus et al., 2016). Hydrological models simulate the processes within the catchment, while hydrodynamic models provide a detailed description of the processes within the water source. The output of the hydrological model, for example fluxes of water and microbes, can be used as input for the hydrodynamic model, in order to represent processes both within the catchment and the water source (Bougeard et al., 2011; De Brauwere et al., 2014a; Sokolova et al., 2018).

The aim of this study was to use hydrological and hydrodynamic models to describe the potential risk of exposing grazing animals to faecal pollutants that may be present in natural drinking water sources on their pastures, using three zoonotic faecal pathogens as model microbes and fictitious pastures in Sweden as examples.

Material and methods

The study assesses two main aspects: (i) microbial contamination by manure from fertilisation or grazing,

modelled by a hydrological model and a hydrodynamic model, and (ii) microbial contamination from human wastewater, modelled by using both models in a backward process (from end-point to source). The hydrological model, HYdrological Predictions for the Environment (HYPE), was used to simulate the pathogen dynamics in the rivers Skeboån and Svartån, as well as the tributaries to Lake Vombsjön: Björkaån, Torpsbäcken and Borstbäcken. The pathogen dynamics in Lake Vombsjön were simulated by the hydrodynamic model MIKE 3 FM, using the HYPE results for the tributaries as input. The geographical points for estimation of pathogen concentrations are shown in Figure 1.

Pathogens

The three faecal pathogens *Salmonella* spp. (salmonella), verotoxin-producing *Escherichia coli* O157:H7 (VTEC) and *Cryptosporidium parvum* (*C. parvum*) were chosen as model organisms. They can all be shed in faeces from infected people and cattle and spread via water.

Salmonella bacteria represent one of the major global causes of human diarrhoeal disease (WHO, 2018). There are more than 2500 serotypes of the main subspecies *Salmonella enterica* subsp. *enterica*. People and animals are usually infected via consumption of contaminated food/feed or water. Infected individuals are often asymptomatic, but clinical infection with enteric as well as systemic symptoms are seen in both animals and humans. Owing to a long-standing national control programme in livestock, including mandatory notification and on-farm eradication of all detected cases, the prevalence of salmonella in Swedish animals is very low (Lahti *et al.*, 2010; Ågren *et al.*, 2016). As a result of this, the level of environmental contamination by salmonella from animals is also low.

VTEC may cause serious enteric disease in humans, but infected cattle are asymptomatic carriers of the bacteria (Karmali *et al.*, 2010). People are mainly infected via contaminated food of animal origin, but outbreaks linked to vegetables contaminated by water are also reported. Outbreaks linked to cattle have occurred in Sweden, most cases occur from July to September (SVA, 2017). In Sweden, detection of infected animals is notifiable if linked to human cases, or if VTEC of clade 8 is identified (SVA, 2017).

This study also comprises the zoonotic cryptosporidium *C. parvum*, while species that are specific for human infections are not included. *C. parvum* causes enteric infection in people and young calves (<1 month). Young calves may shed large amounts of the parasites in their faeces, while older, subclinically infected animals shed negligible concentrations (Constable *et al.*, 2017). *C. parvum* is endemic in Swedish cattle herds, at varying prevalence in different regions and different production systems (Silverlås *et al.*, 2009; Björkman *et al.*, 2015). In infected herds, almost all young calves become infected. Although cryptosporidiosis in humans is notifiable, the proportion of human cases caused by *C. parvum* is unknown, as identification to species level is rarely performed.

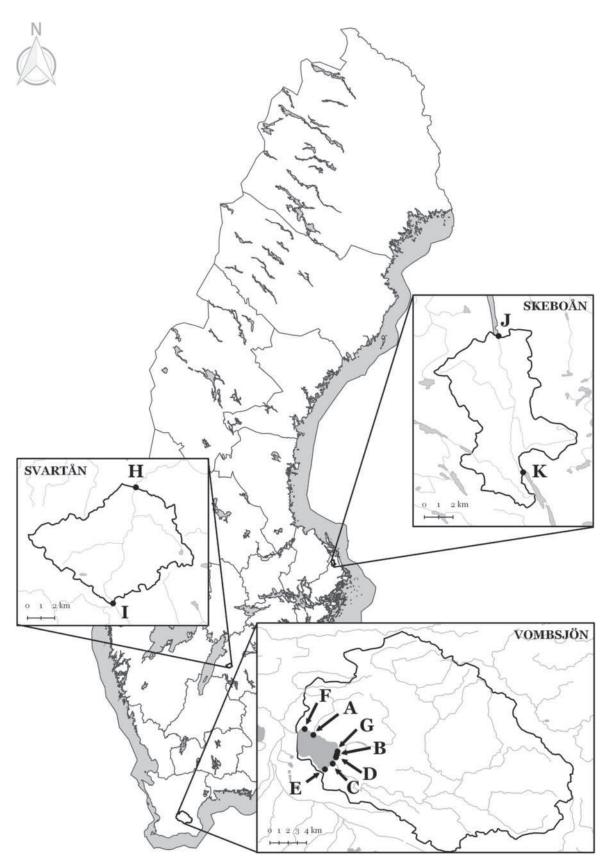


Figure 1 Study areas for hydrological and hydrodynamic modelling. The outlined areas indicate catchment areas with simulated pathogen application on arable land or pastureland. For Lake Vombsjön, A to C indicate the inflow from the tributaries where outputs from the hydrological model also serve as input to the hydrodynamic model. D and E indicate the fictitious drinking water for grazing cattle (i.e. outputs from the hydrodynamic model). F and G indicate the fictitious release points of human wastewater. For Svartån, H indicates the location of the downstream fictitious drinking water point for grazing cattle and I indicates the fictitious release point of human wastewater. In Skeboån, J indicates the location of the downstream fictitious drinking water point for grazing cattle and K indicates the fictitious release point of human wastewater.

Study areas

The fictitious pastures for estimation of risks to grazing cattle were located adjacent to Lake Vombsjön, the rivers Skeboån and Svartån in Sweden. The lake was chosen based on location (agricultural area and surrounded by arable land and pastureland) and the fact that it was also subject to other studies in collaborating projects. Skeboån was also included in some collaborating projects, and Svartån had previously been implicated in salmonella outbreaks in cattle. The water sources and water catchment areas are shown in Figure 1. This figure also shows the fictitious points for wastewater release and cattle exposure on fictitious pastures.

Model inputs

Animal sources. Contamination from infected animals was simulated in two pathways: application of manure on arable land (fertilisation scenario) or manure from grazing animals on pasture (grazing scenario), as described in Sokolova *et al.* (2018). In short, the amounts of the pathogens salmonella, VTEC and *C. parvum* on arable land and pastures were estimated based on calculations of theoretical pathogen concentration in the faeces from infected animals, herd prevalence and within-herd prevalence (see Supplementary material S1). As detailed data could not be obtained, the concentrations in faeces were estimated by expert judgement based on published data. The estimated pathogen numbers were set to a worst-case scenario with two levels ('medium' and 'high'), assuming endemic infection with high between-herd and within-herd prevalence (Sokolova *et al.*, 2018, Supplementary material S1).

In the fertilisation scenario, half of the yearly allowed manure load was assumed to be applied on all arable land in the catchment areas on 8 May 2012, before an event with high observed rainfall on 10 May 2012. This should be considered as a worst-case scenario. It was assumed that 90% of the manure originated from cattle, no reduction in pathogen content occurred during manure storage and before the rainfall 95% of the manure was ploughed into the soil, with 5% remaining on the soil surface.

In the grazing scenario, contaminated manure from the maximum number of cattle allowed (7 per hectare) was applied on all pastureland in the catchment areas during a 14-day period beginning on 10 June 2012, followed by an event with high observed rainfall on 24 June 2012. It was assumed that all pastureland in the area was grazed by cattle, 50% from dairy herds and 50% from beef herds, and that the number of beef calves below 2 months of age was 0.5 per hectare on 50% of the area (beef calves are the only calves that graze on pasture in Sweden). Again, this maximum load is a worst-case scenario.

Human sources. Contamination from human wastewater discharge was simulated by backwards modelling, estimating the number of pathogens required to be released from a fictitious wastewater discharge (see Figure 1) to reach a concentration high enough to cause infection in cattle drinking water at the points on the fictitious pastures (Figure 1). A set of simulations with fictitious release of

wastewater were performed to assess the effect of hydrometeorological conditions on the concentration of pathogens. For the lake, the simulations were performed for the conditions of different wind directions over the lake. For the rivers, the simulations were performed for the conditions of different stream flow magnitudes of the receiving river.

To put the results of the backwards modelling into perspective, reported data were used to calculate the potential pathogen concentrations in untreated wastewater from a fictitious Swedish wastewater treatment plant. For salmonella and VTEC, these calculations assumed 25 000 people living in the households connected to the wastewater treatment plant and used national incidence data, reported shedding durations and concentrations (Schönning et al., 2007; Sundström, 2010; SVA, 2017). For *C. parvum*, human incidence data could not be estimated (as the causative agent for human cryptosporidiosis is not reported on species level), but published data on concentrations in untreated wastewater were available and therefore used (Ottoson et al., 2006; Public Health Agency, 2011). It was assumed that pathogen release occurred due to a single 24-h overflow in the wastewater treatment plant. Details are available as Supplementary material S1.

Water quality modelling

The fate and transport of pathogens within the study areas were simulated using process-based deterministic water quality modelling. The processes in the catchments of the studied rivers Skeboån and Svartån and of the tributaries of Lake Vombsjön were simulated using the hydrological model HYPE, while the circulation within Lake Vombsjön was simulated using the hydrodynamic model MIKE 3 FM. The hydrological and hydrodynamic models were validated by comparing the modelling results with observed discharge and water level data, respectively. Details are available in Sokolova et al. (2018) and as Supplementary material S2. HYPE was chosen because it is already set up to simulate high-resolution fluxes of water, nitrogen and phosphorus for the whole of Sweden, and it considers the hydrological response of several different land uses and soil types. The MIKE 3 FM model was chosen because it has been previously successfully applied to describe hydrodynamic conditions and contaminant fate and transport in lakes (Sokolova et al., 2013).

Hydrological model. The HYPE (Lindström et al., 2010; http://hypecode.smhi.se/) model used simulates water and substances in soils, rivers and lakes. In addition to pure hydrological studies, the model has mainly been used for assessments of nutrients in surface water (Strömqvist et al., 2012; Yin et al., 2016). The use of HYPE for simulating the fate of pathogens in the environment has been described by Sokolova et al. (2018).

Land-based sources of pathogens in the model are either livestock manure applied to the land or manure from grazing animals. Pathogens are released from the source when exposed to rainfall and snowmelt. They may either infiltrate into the top soil or enter the stream network directly via surface runoff depending on water flow pathways. Pathogens

infiltrating into the soil may, to a various degree, sorb to soil particles. The number of pathogens adsorbing or desorbing is driven by the difference between the concentration in soil water and an equilibrium concentration. Pathogens in the soil water may reach the stream via soil drainage. Pathogens discharged into the stream are routed with the water through the model's network of streams and lakes. Exponential decay of pathogens is simulated in all model compartments, that is in the source, soil (both adsorbed and in the soil solute), lakes and rivers. Half-lives in the order of days or weeks are used for the pathogens simulated in this study. In reality, decay will be the sum of microbial die-off and growth, and depends on, for example, temperature and nutrient availability. Pathogens from point sources may be added to the surface water system. Input from point sources can be either constant during the simulation period or vary in time. A detailed description is found in Sokolova et al. (2018).

Hydrodynamic modelling. To simulate the water flows in the lake, the three-dimensional time-dependent hydrodynamic model MIKE 3 FM (MIKE Powered by DHI) was used. The model consists of continuity, momentum, temperature, salinity and density equations, and is closed using a turbulent closure scheme. The application of this model to Lake Vombsjön has been described by Sokolova et al. (2018). The model was set up to account for the hydrometeorological conditions (e.g. wind and precipitation on the lake surface), and to simulate the heat exchange between the atmosphere and the lake. The water density was formulated as a function of temperature. In order to simulate the pathogen fate and transport in Lake Vombsjön, the microbial water quality model ECO Lab was coupled to the hydrodynamic model of the lake. ECO Lab used flow fields from the hydrodynamic model to calculate the pathogen concentrations in the lake. The pathogen decay in the lake was described in the same way as in the HYPE model. Sedimentation of *C. parvum* oocysts in the lake was taken into

account. It was conservatively assumed that the oocysts released into the lake were not attached to particles, thus the sedimentation velocity was specified as 0.03 m/day, which is the value suggested for free oocysts (Medema *et al.*, 1998). It was assumed that resuspension does not occur.

Model outputs

The points where pathogen concentrations were estimated are shown in Figure 1. The models give a time series of pathogen concentration at these points, and the peak concentrations (maximum) were used as outputs. Assuming fictitious pastures with grazing cattle at these points, the estimated pathogen concentrations were compared to the estimated concentrations needed for infection of grazing cattle. The latter estimates were calculated based on reported infectious doses for salmonella (Aceto et al., 2011), VTEC (Cray and Moon, 1995; Besser et al., 2002) and C. parvum (Zambriski et al., 2013) and assuming a daily consumption of 50 l of water for adult animals and 10 l for calves. Based on these calculations (for details see Supplementary material S1), the estimated pathogen concentrations required for infection of cattle were: 200 salmonella bacteria per litre for both adult cattle and calves: 200 000 and 30 VTEC bacteria per litre for adult cattle and calves, respectively; and 0.6 C. parvum oocysts per litre for calves.

Results

Animal sources

The maximum pathogen concentrations in the rivers (Skeboån and Svartån) and the tributaries to Lake Vombsjön (Björkaån, Torpsbäcken and Borstbäcken), as simulated by the HYPE model for the different scenarios, are shown in Table 1. For salmonella, none of the simulated concentrations exceeded the levels needed to infect grazing cattle. For VTEC, all the simulated concentrations in the tributaries to

Table 1 Simulated maximum concentrations (pathogen number/litre) in the tributaries to Lake Vombsjön (Björkaån, Torpsbäcken, Borstbäcken) and the rivers Skeboån and Svartån, based on hydrological modelling of worst-case scenarios of pathogen application to arable land and pastureland. Concentrations that could potentially exceed the infectious dose for grazing calves (up to 2 months old) are in bold. Letters A-J indicate the geographic location shown in Figure 1.

Scenario ^a	Björkaån (C)	Torpsbäcken (B)	Borstbäcken (A)	Skeboån (J)	Svartån (H)
Salmonella, fert, medium load	0.06	0.19	0.15	0.02	<0.01
Salmonella, fert, high load	0.14	0.45	0.36	0.05	< 0.01
Salmonella, grazing, medium load	0.08	0.22	0.20	< 0.01	< 0.01
Salmonella, grazing, high load	0.18	0.52	0.46	0.02	< 0.01
VTECb, fert, medium load	83	261	209	29.7	1.6
VTEC, fert, high load	147	460	369	52.5	2.9
VTEC, grazing, medium load	107	300	266	8.8	1.6
VTEC, grazing, high load	188	529	470	15.5	2.7
C. parvum, fert, medium load	60	75	60	7,0	0,4
C. parvum, fert, high load	120	148	119	13.8	0.8
C. parvum, grazing, medium load	122	108	96	4.8	0.7
C. parvum, grazing, high load	241	213	190	9.6	1.4

^aScenarios include salmonella, VTEC and *C. parvum* applied by fertilisation (fert) of arable land with contaminated manure or infected cattle grazing pastureland in the catchment areas. Medium and high refer to the input load in two worst-case scenarios.

bVTEC = verotoxin-producing *Escherichia coli* O157:H7.

Lake Vombsjön exceeded the concentration needed to infect calves. For *C. parvum*, almost all the simulated concentrations, in the tributaries as well as in the rivers, exceeded the concentration needed to infect calves.

The maximum pathogen concentrations at the two fictitious points for grazing animals by Lake Vombsjön, as simulated by the hydrodynamic model, are shown in Table 2. For *C. parvum*, the simulated concentration caused by contaminated water from the tributary Björkaån exceeded the concentration needed for infection of calves. For the other pathogens, the simulated concentrations did not exceed the estimated concentrations needed for infection of grazing cattle.

Human sources

The results from the backwards modelling for Lake Vombsjön and backwards calculation of hydrological modelling outputs in the rivers Skeboån and Svartån are shown in Figure 2. The estimated pathogen loads that would have been needed at the release points to achieve infectious doses for cattle at the grazing points (see Figure 1) were mostly above the potential loads of salmonella and VTEC calculated to be present in a 24-h overflow from a medium size Swedish wastewater treatment plant (25 000 people), indicating that such an overflow is unlikely to cause infection with salmonella or VTEC in cattle grazing at the fictitious grazing points. In contrast, the required pathogen loads of *C. parvum* at the release points were below the potential loads of *C. parvum* in a 24-h overflow, indicating that such an overflow could potentially cause infection in calves grazing at the fictitious grazing points.

Discussion

Exposure of grazing animals to pathogens in their drinking water sources poses a risk to animal health. If infected,

grazing animals may represent a source for further contamination and spread of pathogens. Hence, water quality modelling should not be restricted to human drinking water sources. The backwards modelling approach used in this study is novel and provides insight into the potential risks to animals from wastewater discharges. The wastewater discharge in this study, a single 24-h overflow of untreated wastewater, is not entirely realistic. This discharge was chosen as an initial worst-case scenario that could occur in extreme cases, but not to be expected on a regular basis. If the discharge period is reduced to, for example, 2.4 h, then the load is reduced by 1 Log_{10} . For our scenario, this would mean that the salmonella concentration would be below what would be needed to infect grazing animals, while the risk of *C. parvum* infection in susceptible age categories would remain high even with a shorter discharge period. For a discharge of treated wastewater, the load is reduced according to the efficacy of wastewater treatment. According to a review by McMinn et al. (2017), the median reduction for bacteria is about 2.5 Log₁₀, while in a study by Ottoson et al. (2006), the mean cyst reduction was 2.6 Log₁₀. On the other hand, there are many reports of wastewater effluents containing large numbers of, for example, salmonella (El Boulani et al., 2017).

Most estimates in this study assume a worst-case scenario. It is unlikely that such high pathogen loads would be obtained in the Swedish context, as the overall prevalence of salmonella and VTEC in livestock is far below the assumed herd prevalence in the scenarios. It is, however, not certain that prevalence will stay at such low levels in all regions, unless re-circulation via water is prevented and animal disease control programmes remain in place. Controlling zoonotic infections at herd level is still the most sustainable strategy for prevention of environmental contamination and subsequent human exposure.

Table 2 Simulated maximum concentrations (pathogen number/litre) at each estimation point in Lake Vombsjön (see Figure 1), based on hydrodynamic modelling of contamination from the tributaries (Björkaån, Torpsbäcken, Borstbäcken). Concentrations that could potentially exceed the infectious dose for grazing calves (up to 2 months old) are in bold.

	Point E			Point D		
Scenario ^a	Björkaån (C)	Torpsbäcken (B)	Borstbäcken (A)	Björkaån (C)	Torpsbäcken (B)	Borstbäcken (A)
Salmonella, fert, medium	< 0.01	<0.01	<0.01	< 0.01	<0.01	<0.01
Salmonella, fert, high	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
Salmonella, grazing, medium	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
Salmonella, grazing, high	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
VTEC ^b , fert, medium	0.11	0.03	0.01	3.08	0.03	0.01
VTEC, fert, high	0.20	0.06	0.02	5.44	0.05	0.01
VTEC, grazing, medium	0.10	0.02	0.01	3.23	0.07	0.01
VTEC, grazing, high	0.18	0.03	0.02	5.71	0.12	0.01
C. parvum, fert, medium	0.28	0.03	0.01	2.78	0.07	0.01
C. parvum, fert, high	0.54	0.06	0.02	5.50	0.14	0.02
C. parvum, grazing, medium	0.96	006	0.02	5.76	0.18	0.02
C. parvum, grazing, high	1.91	0.11	0.04	11.40	0.37	0.04

^aScenarios include salmonella, VTEC and *C. parvum* applied by fertilisation (fert) of arable land with contaminated manure or infected cattle grazing pastureland in the catchment areas. Medium and high refer to the input load in two worst-case scenarios.

bVTEC = verotoxin-producing Escherichia coli 0157:H7.

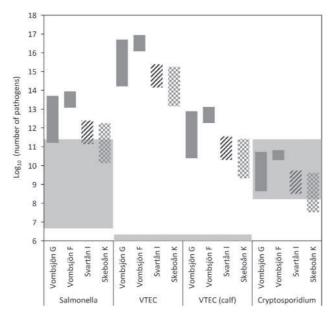


Figure 2 Number of pathogens that could, if released at each simulated point (F and G in Lake Vombsjön, I in river Svartån and K in river Skeboån) result in infection of grazing animals at point D in Vombsjön (full bars), point H in Svartån (striped bar) and point J in Skeboån (chequered bar). The range in the number of pathogens was obtained by studying the fictitious release of wastewater in a set of simulations to assess the effect of hydro-meteorological conditions. For the lake, the simulations were performed for the conditions of different wind directions over the lake. For the rivers, the simulations were performed for the conditions of different stream flow magnitudes of the receiving river. For geographic locations, see Figure 1. Pale grey squares show estimated pathogen loads that could be released during one 24-h overflow of untreated wastewater from a medium size Swedish wastewater treatment plant (serving 25 000 people). VTEC = verotoxin-producing *Escherichia coli* O157:H7

In all modelling, theoretical outputs rely on valid and highresolution input data. In this study, a lot of assumptions had to be made due to lack of data. Estimates based on expert opinion are expected to be biased based on the choice of experts (area of expertise, experience, culture and geography) as well as phenomena such as anchoring (deviation towards the most likely value). Estimates based on literature data may also be biased due to geographic location of the study, study design, etc., and therefore not fully applicable. In addition, the shedding and presence of faecal pathogens would be clustered in space and time (on animal, herd and regional level), and not, as assumed in the scenarios, evenly distributed in all manure applied onto the land. Further scenarios to account for this would have made the results more complicated and less useful, but this limits the direct use of the results as regards the output figures per se. In order to run the models, points for pathogen release and uptake by grazing cattle had to be specified. This affects the ability to draw general conclusions, as the results pertain to these specific locations. Moreover, no model can predict a real-life outcome in detail and this should be kept in mind when using outputs for risk assessment. The main value of the modelling approach lies in the possibilities to adjust inputs and assess the effect of various potential interventions.

No classic validation of the HYPE model's ability to simulate pathogens has been carried out due to lack of data. The plausibility of the model is, however, corroborated by several studies. The flow model is extensively validated both in terms of river flow and also internal variables, such as ground water levels, evapotranspiration and snow depth. The partitioning of runoff into different flow pathway is very important for the runoff of pathogens from land to watercourses. Supporting evidence that the relative importance of different soil flow pathways can be accurately simulated with the HYPE model is given by the model's ability to simulate nitrogen, phosphorus and oxygen-18 (¹⁸O). In addition, the simulation results and sensitivity analysis in Sokolova *et al.* (2018) also indicate that the model behaves in an expected way.

The potential for infection of grazing animals with faecal pathogens has implications for keeping animals on pastures with access to natural water sources. Fences to keep the animals away from water that may be contaminated by, for example, wastewater discharges could be a costly way to manage such risks. As demonstrated in this study, the potential risk needs to be assessed for each location. Backwards modelling could be used for estimation of the risk and how this can be changed by introducing various interventions. Such interventions may include adjusting the time periods when cattle are allowed to graze and/or when manure is applied to different patches of arable land based on weather forecasts, as well as reducing the frequency and magnitude of wastewater discharges. The cost-efficiency of different strategies to reduce the risks will depend on local circumstances.

Although the figures in the results of this study should not be taken *ad notam*, it is clear that if grazing animals are exposed to faecal pathogens in natural water sources, the infectious dose for most pathogens is more easily reached for calves than for adult animals. As young calves are also the main shedders of *C. parvum* in infected herds, it seems like a good general advice to avoid putting young calves on pastures adjacent to such water sources.

Acknowledgements

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Declaration of interest

The authors declare no competing interest.

Ethics statement

No ethical approval was necessary as the study contains only theoretical and/or publicly available data.

Software and data repository resources

Information on the hydrological model set-up and flow data are available at https://www.smhi.se/klimatdata/hydrologi/vattenwebb. No other data are deposited in an official repository.

Supplementary materials

To view supplementary material for this article, please visit https://doi.org/10.1017/S1751731118003415

References

Aceto H, Miller SA and Smith G 2011. Onset of diarrhea and pyrexia and time to detection of *Salmonella enterica* subsp enterica in feces in experimental studies of cattle, horses, goats, and sheep after infection per os. Journal of the American Veterinary Medical Association 238, 1333–1339.

Ågren ECC, Sternberg Lewerin S, Wahlström H, Emanuelson U and Frössling J 2016. Low prevalence of *Salmonella* in Swedish dairy herds highlight differences between serotypes. Preventive Veterinary Medicine 125, 38–45.

Besser TE, Richards BL, Rice DH and Hancock DD 2002. *Escherichia coli* 0157:H7 infection of calves: infectious dose and direct contact transmission. Epidemiology and Infection 127, 555–60.

Björkman C, Lindstrom L, Oweson C, Ahola H, Troell K and Axen C 2015. Cryptosporidium infections in suckler herd beef calves. Parasitology 142, 1108–1114.

Bougeard M, Le Saux JC, Pérenne N, Baffaut C, Robin M and Pommepuy M 2011. Modeling of *Escherichia coli* fluxes on a catchment and the impact on coastal water and shellfish quality. Journal of the American Water Resource Association 47, 350–366.

Bradford SA, Morales VL, Zhang W, Harvey RW, Packman Al, Mohanram A and Welty C 2013. Transport and fate of microbial pathogens in agricultural settings. Critical Reviews in Environmental Science and Technology 43, 775–893.

Cho KH, Pachepsky YA, Oliver DM, Muirhead RW, Park Y, Quilliam RS and Shelton DR 2016. Modeling fate and transport of fecally-derived microorganisms at the watershed scale: State of the science and future opportunities. Water Research 100, 38–56.

Clark RG, Fenwick SG, Nicol CM, Marchant RM, Swanney S, Gill JM, Holmes JD, Leyland M and Davies PR 2004. Salmonella Brandenburg – emergence of a new strain affecting stock and humans in the South Island of New Zealand. New Zealand Veterinary Journal 52, 26–36.

Constable PD, Hinchcliff KW, Done SH and Grünberg W (Editors) 2017. Veterinary medicine: A textbook of the diseases of cattle, horses, sheep, pigs and goats, 11th edition. Elsevier Ltd, St Louis, MO, USA.

Cray WC Jr and Moon HW 1995. Experimental infection of calves and adult cattle with *Escherichia coli* O157:H7. Applied and Environmental Microbiology 61, 1586–1590.

De Brauwere A, Gourgue O, de Brye B, Servais P, Ouattara NK and Deleersnijder E 2014a. Integrated modelling of faecal contamination in a densely populated river-sea continuum (Scheldt River and Estuary). Science of the Total Environment 468-469, 31–45.

De Brauwere A, Ouattara NK and Servais P 2014b. Modeling fecal indicator bacteria concentrations in natural surface waters: a review. Critical Reviews in Environmental Science and Technology 44, 2380–2453.

Dienus O, Sokolova E, Nyström F, Matussek A, Löfgren S, Blom L, Pettersson TJR and Lindgren P-E 2016. Norovirus dynamics in wastewater discharges and in the recipient drinking water source: long-term monitoring and hydrodynamic modeling. Environmental Science and Technology 50, 10851–10858.

El Boulani A, Mimouni R, Mannas H, Hamadi F and Chaouqy N 2017. Salmonella in wastewater: identification, antibiotic resistance and the impact on the marine environment. In: Current topics in Salmonella and Salmonellosis (ed. M Mares). IntechOpen, London, United Kingdom https://doi.org/10.5772/67298.

Jamieson R, Gordon R, Joy D and Lee H 2004. Assessing microbial pollution of rural surface waters: a review of current watershed scale modeling approaches. Agricultural Water Management 70, 1–17.

Karmali MA, Gannon V and Sargeant JM 2010. Verocytotoxin-producing *Escherichia coli* (VTEC). Veterinary Microbiology 140, 360–370.

Lahti E, Ivarsson S, Ågren E, Thelander M, Sjöland L, Eriksson E, Aspan A, Szanto E, Löfdahl S, Plym Forshell L and Wahlström H 2010. A prolonged outbreak of Salmonella. Reading affecting humans and animals in Sweden. Proceedings of I3S International Symposium on Salmonella and Salmonellosis, 21–28 June 2010, St Malo, France, pp. 333–335.

Lindström G, Pers CP, Rosberg R, Strömqvist J and Arheimer B 2010. Development and test of the HYPE (Hydrological Predictions for the Environment) model

 a water quality model for different spatial scales. Hydrology Research 41, 295–319.

McMinn BR, Ashbolt NJ and Korajkic A 2017. Bacteriophages as indicators of faecal pollution and enteric virus removal. Letters in Applied Microbiology 65, 11–26.

Medema GJ, Schets FM, Teunis PFM and Havelaar AH 1998. Sedimentation of free and attached Cryptosporidium oocysts and Giardia cysts in water. Applied Environmental Microbiology 64, 4460–4466.

Oliver DM, Porter KDH, Pachepsky YA, Muirhead RW, Reaney SM, Coffey R, Kay D, Milledge DG, Hong E, Anthony SG, Page T, Bloodworth JW, Mellander P-E, Carbonneau PE, McGrane SJ and Quilliam RS 2016. Predicting microbial water quality with models: over-arching questions for managing risk in agricultural catchments. Science of the Total Environment 544, 39–47.

Ottoson J, Hansen A, Westrell T, Johansen K, Norder H and Stenström TA 2006. Removal of noro- and enteroviruses, Giardia cysts, Cryptosporidium oocysts, and fecal indicators at four secondary wastewater treatment plants in Sweden. Water Environment Research 78, 828–834.

Public Health Agency of Sweden 2011. Cryptosporidium i Östersund: Smittskyddsinstitutets arbete med det dricksvattenburna utbrottet i Östersund 2010–2011 (in Swedish). Retrieved on 20 April 2018 from https://www.folkhalsomyndigheten.se/contentassets/6ba0208adacc460b8aa203fadea39292/cryptosporidium-i-ostersund.pdf.

Schönning C, Westrell T, Stenström TA, Arnbjerg-Nielsen K, Hasling AB, Høibye L and Carlsen A 2007. Microbial risk assessment of local handling and use of human faeces. Journal of Water and Health 5, 117–128.

Silverlås C, Emanuelson U, de Verdier K and Björkman C 2009. Prevalence and associated management factors of Cryptosporidium shedding in 50 Swedish dairy herds. Preventive Veterinary Medicine 90, 242–253.

Sokolova E, Lindström G, Pers C, Strömqvist J, Sternberg Lewerin S, Wahlström H and Sörén K 2018. Water quality modelling: microbial risks associated with manure on pasture and arable land. Journal of Water and Health 16, 549–561.

Sokolova E, Pettersson TJR, Bergstedt O and Hermansson M 2013. Hydrodynamic modelling of the microbial water quality in a drinking water source as input for risk reduction management. Journal of Hydrology 497, 15–23.

Strömqvist J, Arheimer B, Dahné J, Donnelly C and Lindström G 2012. Water and nutrient predictions in ungauged basins – set-up and evaluation of a model at the national scale. Hydrological Sciences Journal 57, 229–247.

Sundström K 2010. Samhällskostnader för salmonellos, campylobacterios och EHEC. Bilaga 9 Betänkande Folkhälsa – Djurhälsa: Ny ansvarsfördelning mellan stat och näring. (SOU 2010:106, in Swedish). Retrieved on 20 April 2018 from http://www.regeringen.se/49bbab/contentassets/85bc16894e354a5-ba40187238673aa51/folkhalsa—djurhalsa-ny-ansvarsfordelning-mellan-stat-och-naring-del-c-bilaga-9-sou-2010106.

SVA 2017. Surveillance of infectious diseases in animals and humans in Sweden 2016, National Veterinary Institute (SVA), Uppsala, Sweden. Retrieved on 20 April 2018 from http://www.sva.se/globalassets/redesign2011/pdf/om_sva/publikationer/surveillance-2016-w.pdf.

Vanselow BA, Hum S, Hornitzky MA, Eamens GJ and Quinn K 2007. Salmonella typhimurium persistence in a Hunter Valley dairy herd. Australian Veterinary Journal 85, 446–450.

WHO 2018. Non-typhoidal salmonella. Retrieved on 20 April 2018 from http://www.who.int/mediacentre/factsheets/fs139/en/.

Yin Y, Jiang S, Pers C, Yang X, Liu Q, Yuan J, Yao M, He Y, Luo X and Zheng Z 2016. Assessment of the spatial and temporal variations of water quality for agricultural lands with crop rotation in China by using a HYPE model. International Journal of Environmental Research and Public Health 13, 336.

Zambriski JA, Nydam DV, Wilcox ZJ, Bowman DD, Mohammed HO and Liotta JL 2013. *Cryptosporidium parvum*: determination of ID_{50} and the dose-response relationship in experimentally challenged dairy calves. Veterinary Parasitology 197, 104–112.